## Chemical and Cytotoxic Constituents from the Stem of Machilus zuihoensis

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Five new compounds, including a novel lactone, machilactone (=rel-(2R,3aR,6E,6aS)-2-heptadecyl-3amethyl-6-octadecylidene-6,6a-dihydrofuro[2,3-d][1,3]dioxol-5(3aH)-one; **1**), a new sesquiterpene, 3,4-dihydroxy- $\beta$ -bisabolol (=rel-(1R,2S,4R)-1-[(1R)-1,5-dimethylhex-4-enyl]-1-methylcyclohexane-1,2,4-triol; **2**), a new secobutyrolactone, methyl (2E)-2-(1-hydroxy-2-oxopropyl)eicos-2-enoate (**3**), two new butyrolactones, machicolide A (**4**) and machicolide B (**5**) (=3E,4R,5R)- and (3Z,4R,5R)-4,5-dihydro-4-hydroxy-5-methoxy-5methyl-3-octadecylidenefuran-2(3H)-one, resp.) as a mixture, together with known caryophyllene oxide (=4,12,12-trimethyl-9-methylene-5-oxatricyclo[8.2.0.0<sup>4,6</sup>]dodecane), hexacosane, tetracosanoic acid, isomahubanolide-23 (=(3E,4R)-4,5-dihydro-4-hydroxy-5-methylidene-3-octadecylidenefuran-2(3H)-one), and  $\beta$ -bisabolol (=(1S)-1-[(1S)-1,5-dimethylhex-4-enyl]-4-methylcyclohex-3-en-1-ol) were isolated from the stem wood of *Machilus zuihoensis*. The structures of these compounds were established by spectroscopic studies. The eicos-2-enoate (**3**) and  $\beta$ -bisabolol exhibited marginal cytotoxicity against NUGC and HONE-1 cancer cell lines *in vitro*.

**Introduction.** – Recently, studies on Lauraceous plants of chemical interest have gradually brought cytotoxic constituents into focus [1-7]. *Machilus zuihoensis* HAYATA (Lauraceae), an endemic species in Taiwan, is an evergreen medium-sized tree found widely throughout the island from the lowlands up to 1500 m [8]. Its bark is an incense material for joss sticks. Earlier investigations have established the presence of the two alkaloids: (-)-L-N-norarmepavine and  $(\pm)$ -N-norarmepavine in the stem [9], and of four neolignans in the bark of this plant [10]. Of this plant, only the stem wood showed significant cytotoxicity on high-throughput screening against NUGC-3 and HONE-1 cancer cell lines *in vitro*. Examination of the hexane-soluble part of the stem wood led to the isolation of five new compounds: machilactone (1), 3,4-dihydroxy- $\beta$ -bisabolol (2), methyl (2*E*)-2-(1-hydroxy-2-oxopropyl)eicos-2-enoate (3), and the mixture of machicolide A (4) and machicolide B (5). We now report the isolation and characterization of 1–5 by spectral analyses and the cytotoxicity of the isolates.

**Results and Discussion.** The optically active machilactone (1), obtained as colorless needles, was analyzed for  $C_{41}H_{76}O_4$  from its FAB-HR-MS (633.5803 ( $[M + H]^+$ )). The UV spectrum (243 nm) and IR data (1763 and 1677 cm<sup>-1</sup>) indicated the presence of an  $\alpha$ -alkylidene-substituted  $\gamma$ -lactone skeleton and the <sup>1</sup>H- and <sup>13</sup>C-NMR spectra suggested the presence of a  $\gamma$ -butyrolactone moiety [4] connected to another aliphatic chain by an acetal linkage.



The <sup>1</sup>H-NMR signals of **1** at  $\delta$  7.09 and 2.40 were assignable to CH=C(6) and  $CH_2CH=C(6)$  based on HMQC and <sup>1</sup>H,<sup>1</sup>H-COSY experiments. Comparison with available butyrolactones of this kind and a significant fragment at m/z 393 in the FAB-MS suggested the presence of an octadecylidene side chain at C(6). The geometry of the alkylidene moiety was determined to be (*E*) based on the chemical shifts of CH=C(6) ( $\delta$  7.09) and  $CH_2CH=C(6)$  ( $\delta$  2.40) [4]. This was also confirmed by the NOE difference spectrum, which showed the correlation  $H-C(6a)/CH_2CH=C(6)$ . The <sup>1</sup>H-NMR signal at  $\delta$  5.01 was assigned to H-C(2) as this proton correlated with the dioxygenated aliphatic C(2) signal at  $\delta$  103.9 in the HMQC and with the neighboring CH-C(2) ( $\delta$  1.68) in the COSY plot. From the <sup>1</sup>H,<sup>1</sup>H-COSY experiment and the molecular formula of **1**, the length of the saturated aliphatic side-chain portion connected to the exocyclic C=C bond was deduced to be C<sub>17</sub>. A Me group at  $\delta$  1.71 was placed at C(3a), as these protons showed <sup>2</sup>J correlation with C(3a) ( $\delta$  109.8) and <sup>3</sup>J butyrolactone moiety by a O-C-O linkage was confirmed by the <sup>13</sup>C-NMR signals of two aliphatic dioxygenated C-atoms at  $\delta$  103.9 (C(2)), 109.8 (C(3a)) and the correlation observed between H-C(6a) ( $\delta$  4.89) and C(2) in the HMBC plot.

According to HMQC, COSY, NOE, and HMBC data, the structure of **1** was elucidated as 2-heptadecyl-3a-methyl-6-octadecylidene-6,6a-dihydrofuro[2,3-*d*][1,3]di-oxol-5-(3a*H*)-one was attributed to **1**, with the relative *cis* configuration of H-C(6a)/Me-C(3a) and  $H-C(6a)/CH_2-C(2)$  and (6*E*)-configuration.

Compound **2**, obtained as colorless needles, has the molecular formula  $C_{15}H_{28}O_3$ , as determined by FAB-HR-MS. According to the IR, <sup>1</sup>H-NMR, <sup>13</sup>C-NMR, DEPT, COSY, NOE, HMQC, and HMBC studies, the structure of **2** was elucidated as 3,4-dihydroxy- $\beta$ -bisabolol, *i.e.* as *rel-*(1*R*,2*S*-4*R*)-1-[(1*R*)-1,5-dimethylhex-4-enyl]-1-methylcyclohexane-1,2,4-triol.

A strong IR absorption band at 3322 cm<sup>-1</sup> indicated the presence of OH groups in 2. The <sup>13</sup>C-NMR and DEPT experiments showed the 15 C resonances for 4 Me, 5 CH<sub>2</sub>, 3 CH, and 3 quarternary C-atoms, suggesting that 2 could be a sesquiterpene of the  $\alpha$ - or  $\beta$ -bisabolene type. The presence of a Me at  $\delta$  0.93 (J=6.6, Me – C(1')) supported that **2** was of the latter type. A close comparison of <sup>13</sup>C-NMR data with  $\beta$ -bisabolol [11], also isolated in this study (see below), the missing signal for the C=C bond, and the presence of further two oxygenated C-atoms at  $\delta$  71.7 and 74.9 suggested that **2** could be 3,4-dihydroxy- $\beta$ -bisabolol. The <sup>1</sup>H-NMR showed signals for two Me groups at  $\delta$  1.60 and 1.68, assigned to Me – C(6') and Me – C(5'), respectively, as they showed the correlation with each other as well as  ${}^{2}J$  correlation with C(5') and  ${}^{3}J$  correlation with C(4') in its HMBC plot. The third Me group at  $\delta$  0.93 (d, J = 6.6) was attributed to Me – C(1'), and the remaining Me signal at  $\delta$  1.33 to Me-C(1) based on HMBC studies. An olefinic proton at  $\delta$  5.09 was placed at C(4') and another proton with a br. s at  $\delta$  3.52 was assigned to H–C(2). Three OH groups were recognized in the <sup>1</sup>H-NMR at  $\delta$ 1.10, 1.90, and 3.80 by addition of  $D_2O$ . The strong NOE difference correlation between Me – C(1) and H – C(2) supported the relative cis configuration of the dihydroxy groups at C(1) and C(2) (axial position), which was also evidenced by the Me-C(1) signal at  $\delta$ (C) 27.7 [12][13]. The NOE difference spectrum also showed correlation between the H-C(2) and  $CH_2(2')$ , supporting the equatorial position of OH-C(4). Other NOE correlations between Me-C(1') and  $H_b$ -C(3) and  $H_b$ -C(2') were also found.

Compound **3** was obtained as colorless oil. The molecular formula was deduced to be  $C_{24}H_{44}O_4$  by FAB-HR-MS as it showed the  $[M + H]^+$  ion at m/z 397.3318. From the spectral studies, the structure of **3** was elucidated as methyl (2*E*)-2-(1-hydroxy-2-oxopropyl)eicos-2-enoate (**3**), which was further substantiated by COSY, NOESY, and HMQC experiments. The zero  $[a]_D^{20}$  value of **3** suggested that it was considerably racemized.

The IR spectrum of **3** showed absorption bands due to the presence of OH (3462 cm<sup>-1</sup>) and C=O (1726 cm<sup>-1</sup>) groups. The <sup>1</sup>H-NMR spectrum exhibited the signals for an olefinic proton at  $\delta$  7.08 (t, J = 8.0), an oxymethine proton at  $\delta$  4.90 (br. *s*), an OH group at  $\delta$  4.02 (br. *s*) (D<sub>2</sub>O exchangeable), a MeO group at  $\delta$  3.73

(s), an Ac group at  $\delta 2.15$  (s), CH<sub>2</sub> groups at  $\delta 2.35$  (q), 1.51 (m), and 1.26 (br. s), and a terminal Me group at  $\delta 0.88$  (t, J = 6.8), similar to that of the methanolysis product of isolineranolide E [14]. The geometry of the alkene chain of **3** was determined to be the (*E*) form based on the chemical shifts of H–C(3) ( $\delta$  7.08) and H–C(4) ( $\delta$  2.35) [14].

The optically active mixture of machicolide A (4; (*E*); 65%) and machicolide B (5; (*Z*); 35%) was obtained as a colorless oil. The molecular formula was deduced to be  $C_{24}H_{44}O_4$  from FAB-HR-MS (397.3308,  $[M + H]^+$ ). The UV spectrum exhibited the absorption maxima at 216 nm, and the IR absorptions inferred the presence of an OH group (3420 cm<sup>-1</sup>) and an  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone (1746 and 1683 cm<sup>-1</sup>). The data suggested the structure of (3*E*)- and (3*Z*)-4,5-dihydro-4-hydroxy-5-methoxy-5-methyl-3-octadecylidenefuran-2(3*H*)-one for **4** and **5**, respectively.

The <sup>1</sup>H-NMR spectrum of **4** showed signals for an olefinic proton ( $\delta$  6.99 (*td*, J = 8.0, 1.6)), an oxymethine proton ( $\delta$  4.54 (br. *s*)), a MeO group ( $\delta$  3.39 (*s*)), a Me group ( $\delta$  1.60 (*s*)) and CH<sub>2</sub> groups ( $\delta$  2.38 (*m*); 1.50 (*m*); 1.26 (br. *s*)), and a striking resemblance to those of litseakolide F (**6**) [7]. The side chain at the  $\gamma$ -lactone of **4** was found to be an octadecylidene group from its molecular formula. The <sup>1</sup>H-NMR spectrum of **5** showed high similarity to that of litseakolide G (**7**) except for an octadecylidene group in **5**.

The configuration at C(4) in both compounds was reasonably determined to be (4R) [6][9][15], and the NOESY data supported (5*R*) configuration.

The NOESY of 4/5 showed correlations between the H-C(4) and MeO-C(5) signals, like in the case of 6 and 7, supporting the (5*R*) configuration [7].

Besides **1**–**5**, the known caryophyllene oxide (=4,12,12-trimethyl-9-methylene-5oxatricyclo[8.2.0.0<sup>4,6</sup>]dodecane), hexacosane, tetracosanoic acid, isomahubanolide-23 (=(3*E*),4*R*)-4,5-dihydro-4-hydroxy-5-methylidene-3-octadecylidenefuran-2(3*H*)-one), and  $\beta$ -bisabolol (=(1*S*)-1-[(1*S*)-1,5-dimethylhex-4-enyl]-4-methylcyclohex-3-en-1-ol) were also isolated from the stem wood of *Machilus zuihoensis*.

The cytotoxicity activity of all isolates were tested *in vitro* against NUGC-3 and HONE-1 cancer cell lines. The eicos-2-enoate **3** showed marginal activity, with  $ED_{50}$  values of 6.33 and 6.44 µg ml<sup>-1</sup>, respectively, as did  $\beta$ -bisabolol with  $ED_{50}$  values of 6.68 and 6.73 µg ml<sup>-1</sup>, respectively.

## **Experimental Part**

General. TLC: silica gel 60  $F_{254}$  precoated plates (*Merck*). Column chromatography (CC): silica gel 60 (*Merck* 70–230 mesh, 230–400 mesh, ASTM). M.p.: Uncorrected. Optical rotation: Jasco DIP-370 polarimeter; in CHCl<sub>3</sub>. UV Spectra: Jasco UV-240 spectrophotometer;  $\lambda_{max}$  (log  $\varepsilon$ ) in nm. IR Spectra: Perkin-Elmer 2000 FT-IR spectrophotometer;  $\nu$  in cm<sup>-1</sup>, <sup>1</sup>H-, <sup>13</sup>C- and 2D-NMR: Varian Unity-Plus-400 and Jeol GSX-600 spectrometers;  $\delta$  in ppm, J in Hz. EI-MS Spectra: VG-Biotech Quatro-5022 spectrometer; m/z (rel. %). FAB-HR-MS: Jeol SX102A mass spectrometer.

*Plant Material.* The stem wood of *M. zuihoensis* was collected at Mutan, Pingtung County, Taiwan, in January 2000. A voucher specimen (No. Chen 2280) was deposited in the Herbarium of the School of Pharmacy, Kaohsiung Medical University, Kaohsiung, Taiwan, R.O.C.

*Extraction and Isolation.* Shade-dried and powered plant material (23 kg) was extracted with MeOH. The extract (2 kg) was partitioned into hexane (500 g), AcOEt (87 g), and MeOH-soluble (500 g) parts. The hexane fraction (81 g) was submitted to CC (silica gel, hexane/AcOEt step gradients): *Fractions* 1-7. *Fr.* 1 (AcOEt/hexane 1:4, 5 g) was subjected to CC (silica gel; AcOEt/hexane) to give tetracosanoic acid (5 mg). Repeated

purification of *Fr. 3* (AcOEt/hexane 2:3; 4 g) by CC (silica gel; AcOEt/hexane) gave **1** (15 mg), **2** (10 mg), hexacosane (30 mg), isomahubanolide-23 (100 mg), and  $\beta$ -bisabolol (10 mg). *Fr. 4* (AcOEt/hexane 1:1; 100 mg) was subjected to prep. TLC (AcOEt/CH<sub>2</sub>Cl<sub>2</sub> 1:4) to furnish **3** (15 mg) and caryophyllene oxide (20 mg). *Fr. 6* (AcOEt/hexane 4:1; 20 mg) was purified by prep. TLC (Me<sub>2</sub>CO/CHCl<sub>3</sub> 1:1): **4/5** (3 mg).

rel-(2R,3aR,6E,6aS)-2-Heptadecyl-3a-methyl-6-octadecylidene-6,6a-dihydrofuro[2,3-d][1,3]dioxol-5(3aH)one (= Machilactone A; **1**). Colorless needles. M.p. 58–60°.  $[a]_{20}^{20} = +20.95$  (c = 0.082, CHCl<sub>3</sub>). UV (CHCl<sub>3</sub>): 243 (4.36). IR (KBr): 2917, 2846, 1763, 1677, 1468, 1389, 1266, 1116, 1057. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 600 MHz): 0.88 (t, J = 7.2, Me(18'), Me(17'')); 1.26 (br. *s*, 29 CH<sub>2</sub>); 1.50 (m, CH<sub>2</sub>(3')); 1.68 (m, CH<sub>2</sub>(1'')); 1.71 (s, Me–C(3a)); 2.40 (m, CH<sub>2</sub>(2')); 4.89 (d, J = 1.2, H–C(6a)); 5.01 (t, J = 4.8, H–C(2)); 7.09 (td, J = 7.8, 1.8, H–C(1')). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 150 MHz): 14.1 (C(18'), C(17'')); 22.7 (Me–C(3a)); 22.4, 23.6, 26.9, 27.2, 29.3, 29.4, 29.5, 29.6, 29.7, 31.9, 32.0 (29 C, CH<sub>2</sub>); 28.2 (C(3')); 30.2 (C(2')); 32.7 (C(1'')); 78.4 (C(6a)); 103.9 (C(2)); 109.8 (C(3a)); 126.3 (C(6)); 150.8 (C(1')); 168.1 (C(5)). FAB-MS: 633 (26, [M + H]<sup>+</sup>), 569 (12), 525 (20), 481 (16), 437 (20), 393 (28), 365 (71), 242 (47), 205 (100). FAB-HR-MS: 633.5803 (C<sub>41</sub>H<sub>77</sub>O<sub>4</sub><sup>+</sup>; calc. 633.5822).

rel-(*1*R,2S,4R)-*1*-[(*1*R)-*1*,5-Dimethylhex-4-enyl]-1-methylcyclohexane-*1*,2,4-triol (= 3,4-Dihydroxy- $\beta$ -bisabolol; **2**). Colorless needles. M.p. 138–140°.  $[a]_{D}^{20} = -14.9$  (c = 0.112, CHCl<sub>3</sub>). UV (CHCl<sub>3</sub>): 243 (3.57). IR (KBr): 3322 (OH), 2961, 2925, 2360, 1455, 1375, 1204, 1066, 1029, 914, 850. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 600 MHz): 0.93 (d, J = 6.6, Me–C(1')); 1.06 (m, H<sub>b</sub>–C(6)); 1.10 (br. s, OH); 1.33 (s, Me–C(1)); 1.36 (m, H–C(1')); 1.41 (m, H<sub>b</sub>–C(3)); 1.43 (m, H<sub>b</sub>–C(5)); 1.60 (s, Me(6')); 1.61 (m, H<sub>a</sub>–C(6)); 1.68 (s, Me–C(5')); 1.72 (m, H<sub>b</sub>–C(2')); 1.86 (m, H<sub>a</sub>–C(3)); 1.90 (br. s, OH); 1.91 (m, H<sub>b</sub>–C(3')); 1.96 (m, H<sub>a</sub>–C(2')); 2.02 (m, H<sub>a</sub>–C(5)); 2.10 (m, H<sub>a</sub>–C(3')); 3.52 (br. s, H–C(2)); 3.80 (br. s, OH); 5.09 (t, J = 8.2, H–C(4')). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 150 MHz): 13.5 (Me–C(1')); 17.7 (C(6')); 25.7 (Me–C(5')); 26.5 (C(3')); 27.7 (Me–C(1)); 29.1 (C(5)); 29.8 (C(3)); 30.8 (C(6)); 34.0 (C(2')); 43.6 (C(1')); 71.7 (C(1)); 74.9 (C(2)); 75.8 (C(4)); 124.4 (C(4')); 131.8 (C(5')). FAB-MS: 257 (16, [M + H]<sup>+</sup>), 239 (63, [ $M + H - H_2O$ ]<sup>+</sup>), 221 (61), 203 (50), 69 (100). FAB-HR-MS: 257.2128 (C<sub>15</sub>H<sub>29</sub>O<sub>3</sub><sup>+</sup>; calc. 257.2116).

*Methyl* (2E)-2-(1-hydroxy-2-oxopropyl)eicos-2-enoate (**3**).  $[a]_{D}^{20} = 0$  (c = 0.376, CHCl<sub>3</sub>). UV (MeOH): 213 (4.13). IR (KBr): 3462 (OH), 2922, 2853, 1726, 1646, 1466, 1357, 1275, 1152, 1071. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz): 0.88 (t, J = 6.8, Me(20)); 1.26 (br. s, 14 CH<sub>2</sub>); 1.51 (m, CH<sub>2</sub>(5)); 2.15 (s, MeCOO); 2.35 (q, J = 7.61, CH<sub>2</sub>(4)); 3.73 (s, MeO); 4.02 (br. s, OH–C(1')); 4.90 (br. s, H–C(1')); 7.08 (t, J = 8.0, H–C(3)). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz): 14.0 (C(20)); 27.1 (C(3')); 22.6, 24.7, 29.2, 29.3, 29.4, 29.5, 29.7 (14 C, CH<sub>2</sub>); 28.6 (C(5)); 31.9 (C(4)); 51.9 (MeO); 73.3 (C(1')); 129.6 (C(2)); 149.1 (C(3)); 166.5 (C(1)); 206.3 (C(2')). EI-MS: 397 (4,  $[M + H]^+$ ), 365 (2), 353 (21), 321 (14), 293 (10), 284 (5), 275 (3), 241 (3), 115 (38). FAB-HR-MS: 397.3318 (C<sub>24</sub>H<sub>45</sub>O<sub>4</sub><sup>+</sup>; calc. 397.3318).

*Mixture* **4**/5. Colorless oil.  $[a]_{D}^{20} = +70.28$  (c = 0.044, CHCl<sub>3</sub>). UV (MeOH): 216 sh (4.13). IR (KBr): 3420 (OH), 2921, 2851, 1746, 1683, 1466, 1384, 1283, 1199, 1079, 1021. EI-MS: 397 (42,  $[M + H]^+$ ), 396 (5,  $M^+$ ), 365 (24), 347 (3), 322 (4), 304 (3), 275 (2), 205 (2), 164 (13), 153 (77), 140 (100). FAB-HR-MS: 397.3308 (C<sub>24</sub>H<sub>45</sub>O<sub>4</sub><sup>+</sup>; calc. 397.3319).

(3E,4R,5R)-4,5-Dihydro-4-hydroxy-5-methoxy-5-methyl-3-octadecylidenefuran-2(3H)-one (= Machicolide A;**4**). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz): 0.88 (*t*,*J*= 7.0, Me(18')); 1.26 (br.*s*, 14 CH<sub>2</sub>); 1.50 (*m*, CH<sub>2</sub>(3')); 1.60 (*s*, Me-C(5)); 2.38 (*m*, CH<sub>2</sub>(2')); 3.39 (*s*, MeO); 4.54 (br.*s*, H-C(4)); 6.99 (*td*,*J*= 8.0, 1.6, H-C(1')). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz): 14.1 (Me(18')); 16.1 (Me-C(5)); 22.4, 27.2, 28.8, 29.3, 29.4, 29.5, 29.6, 29.8 (14 C, CH<sub>2</sub>); 29.2 (C(3')); 31.9 (C(2')); 50.3 (MeO); 72.6 (C(4)); 109.3 (C(5)); 130.0 (C(2)); 148.4 (C(1')); 168.0 (C(2)).

(3Z, 4R, 5R) - 4, 5-Dihydro-4-hydroxy-5-methoxy-5-methyl-3-octadecylidenefuran-2(3H)-one (= Machicolide B; 5). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz): 0.88 (t, J = 7.0, Me(18')); 1.26 (br. s, 14 CH<sub>2</sub>); 1.51 (m, CH<sub>2</sub>(3')); 1.55 (s, Me-C(5)); 2.76 (m, CH<sub>2</sub>(2')); 3.40 (s, MeO); 4.40 (br. s, H-C(4)); 6.57 (td, J = 8.0, 1.6, H-C(1')). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz): 14.1 (C(18')); 16.3 (Me-C(5)); 22.7, 26.9, 28.4, 29.4, 29.4, 29.5, 29.6, 29.7 (14 C, CH<sub>2</sub>); 29.3 (C(3')); 33.3 (C(2')); 50.5 (MeO); 75.8 (C(4); 108.8 (C(5)); 129.8 (C(3)); 150.3 (C(1')); 167.8 (C(2)).

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